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1. INTRODUCTION

lazione Icona

Life expectancy of patients with HIV infection receiving combined antiretroviral therapy (cART) is similar but not equal to that of uninfected subjects [1]. Actually, the main factor influencing the prognosis of patients with HIV infection is the onset of non-AIDS-defining events (nADE) as: liver diseases, cardiovascular events, renal impairment, cancers, etc [2]. These events have been related either to the life-style of patients, as intravenous drug use or smoking, or to the continuous state of immune activation that is not fully controlled by cART [3-7]. Indeed, immune-reconstitution after therapy (cART) is often either quantitatively or qualitatively incomplete, and after almost 30 years of HIV infection, CD4 count remains the most important predictive factor of clinical progression concerning HIV-AIDS, but not of immune-activation and of non-ADEs [8]. Since non-ADEs are becoming more common, there is an urgent need for surrogate markers that could be widely used in clinical practice. In patients without HIV infection, a CD4/CD8 ratio <1 has been related to immune-senescence and to all cause mortality [9-11].

In naïve HIV-infected patients with a CD4 count >200 cells/uL, a low CD4 percentage and a low CD4/CD8 ratio before starting cART were predictive of the risk of clinical progression [12]. Moreover, in patients treated with cART and with undetectable HIVRNA, CD4/CD8 ratio was independently associated with T-cell activation [13] and with markers of age-associated disease as carotid intima-media thickness (IMT), arterial stiffness, estimated glomerular filtration rate, muscle wasting and sarcopenia [14]. Aim of the present study was to evaluate in the patients of the Icona cohort the probability of reaching a CD4/CD8 ratio >=1 after starting cART and its possible protective role against the onset of serious non-AIDS related events (nADE) or death for any cause.

2. PATIENTS & METHODS

The Icona Foundation Study is a cohort of HIV-infected patients which superseded the original I.Co.N.A. (Italian Cohort of Antiretroviral-Naïve Patients) study, recruiting HIV positive patients when still ART-naïve regardless of the reason. Clinical, therapeutical, laboratory and immuno-virological parameters are measured, on average, every 4 months. All patients signed consent forms to participate in the Icona Foundation Study, locally in each of the participating clinical sites. Inclusion criteria for this specific study were: 1) starting a cART regimen (at least three drugs) from ART-naïve after the 1st January 1997; 2) reaching a confirmed HIV-RNA<80 copies/mL; 3) CD4/ CD8 ratio at undetectability <=0.8. We conducted two analysis: a) to establish the probability of CD4/CD8 normalization and to determine factors associated to this event and b) to establish whether normalization of CD4/CD8 might be related with the clinical progression defined as the occurrence of serious non-AIDS related events (nADE) or death for any causes. Factors associated with normalization

End-point was the normalization of the ratio defined by two consecutive value of CD4/CD8>=1. Follow-up was censored if the subject lost the HIV-RNA undetectability (first of two consecutive HIV-RNA>80 copies/mL) or the subject died or was lost to follow-up. Kaplan Meier method was used to estimate the probability of normalization. The incidence rate of normalization was calculated dividing the number of events over the total number of person year of follow-up (PYFU). Multivariable Poisson regression model was used to analyze factors independently associated with normalization. Fixed covariates were: gender, age, mode of HIV transmission, years from HIV diagnosis, CD4 nadir, log10 HIV-RNA before cART start, HCV-Ab positive, months from start of therapy, undetectable viral load, calendar year of cART start, CD4/CD8 ratio at baseline. Type of antiretroviral regimen administered was analyzed as time-varying covariate.

<u>CD4/CD8 ratio as a predictor of clinical progression</u>

Serious nADE were [15]: (i) any non AIDS-defining malignancy; (ii) severe non-AIDS defining infections (i.e. potentially life-threatening or requiring intravenous antibiotic); (iii) cardiovascular events (documented acute myocardial infarction, coronary disease requiring invasive procedures, congestive hearth failure, stroke); (iv) hepatic events (decompensated cirrhosis, i.e. variceal bleeding, porto-systemic encephalopathy, refractory ascites, hepatorenal syndrome); (v) acute kidney injury (confirmed estimated glomerular filtrate rate [eGFR]<60ml/min using MDRD formula) or kidney failure requiring dialysis or renal transplantation. Follow-up started with the first of two HIV-RNA<80 cp/ml after therapy start and continued until the occurrence of the following events: serious non-AIDS event, death for any causes, loss to follow-up. Multivariable Poisson regression model was used to analyze the association of CD4/C8 ratio with clinical progression, adjusting for main confounders. Two different multivariable models were fitted to investigate if the role of current CD4/CD8 ratio on clinical progression were independent from current CD4: a) containing current CD4; b) including CD4/CD8 ratio too. The contribution of the additional covariates CD4/CD8 ratio was assessed using Akaike's information criterion (AIC). The model who had the best fit was that with a lowest AIC.

Incidence of CD4/CD8 ratio normalization and its role in the onset of non-AIDS related events.

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Our analysis included 3,236 (81.4%) out of 3,973 subjects who reached viral suppression after cART start, and who had a CD4/CD8 ratio <=0.8.

Table 1. General characteristics of the study po	opulation
Male gender, n(%)	2471
Age, median (IQR)	39
Italian nationality, n(%)	2858
Mode of HIV transmission, n(%)	
Heterosexual contacts	1339
Homosexual contacts	1031
IVDU	669
Other/unknown	197
HIV history	
Years of infection, median (IQR)	2.6
Log ₁₀ HIV-RNA pre cART, median (IQR)	4.80
Nadir cd4 cell/mmc, median (IQR)	223
CDC C stage, n(%)	592
Baseline CD4, cell/mmc, median (IQR)	378
Months to reach viral suppression, median	
(IQR)	5.4
HCV Ab, n(%)	
Positive	717
Negative	1913
Unknown	606
HBs Ag, n(%)	
Positive	142
Negative	2431
Unknown	663
CD4/CD8 ratio at baseline, median (IQR)	0.39

4. DISCUSSION

In conclusion, a minority of patients who start cART reaches a CD4/CD8 ratio≥1. Younger patients, those with a higher CD4/CD8 ratio at time of viral suppression and those starting cART more recently and with higher CD4 count have higher possibilities to achieve normalization. The CD4/CD8 ratio was predictive of serious non AIDS event or death independently from CD4 cell count.

REFERENCES

1. Bhaskaran K, et al. Changes in the risk of death after HIV seroconversion compared with mortality in the general population. JAMA 2008;300:51-59. 2. Mocroft A, et al. Serious fatal and nonfatal non-AIDS-defining illnesses in Europe. J Acquir Immune Defic Syndr 2010;55:262-70. 3. Sainz T, et al. The CD4/CD8 ratio as a marker of T-cell activation, senescence and activation-exhaustion int reated HIV-infected children and young adults. AIDS 2013;27:1513-1516. 4. Deeks SG, et al. HIV infection, inflammation, immunosenescence, and aging. Ann Rev Med 2011;62:141-155. 5. Deeks SG, et al. HIV infection, antiretroviral treatment, ageing, and non-AIDS related morbidity. BMJ 2009;338:a3172. 6. Hsue PY, et al. Role of viral replication, antiretroviral therapy, and immunodeficiency in HIV-associated atherosclerosis. AIDS 2009;23:1059-1067. 7. Franceschi C, et al. Inflamm-aging. An evolutionary perspective on immunosenescence. Ann NY Acad Sci 2000;908:244-254. 8. Lucero T, et al. Rate and predictors of non-AIDS events in a cohort of HIV-infected patients with a CD4 T cell count above 500cells/mm3. AIDS res Hum Retroviruses 2013;29:993-999. 9. Hadrup SR, et al. Longitudinal studies of clonally expanded CD8 T cells reveal a repertoire shrinkage predicting mortality and an increased number of dysfunctional cytomegalovirus-specific T cells in the very elderly. J Immunol 2006;176:2645-2653. 10. Wikby A, et al. Age-related changes in immune parameters in very old population of Swedish people: a longitudinal study. Exp Gerontol 1994:29:531-541 11. Wikby A, et al. Changes in CD8 and CD4 lymphocyte subsets, T cell proliferation responses and non-survival in the very old: the Swedish Iongitudinal OCTO-immune study. Mech Ageing Dev 1998;102:187-198. 12. Guiguet M, et al. CD4+ T-cell percentage is an independent predictor of clinical progression in AIDS-free antiretroviral-naïve patients with CD4+ Tcell count >200 cells/mm3. Antiviral Therapy 2009;14:451-457. 13. Serrano-Villar S, et al. The CD4/CD8 ratio in HIV-infected subjects is independently associated with T-cell activation despite long-term viral suppression. J of Infect 2013;66:57-66. 14. Serrano-Villar, et al. The CD4:CD8 ratio is associated with markers of age-associated disease in virally suppressed HIV-infected patients with immunological recovery. HIV Med 2013 15. Lapadula G, et al; ICONA Foundation Study. Risk of clinical progression among patients with immunological nonresponse despite virological suppression after combination antiretroviral treatment. AIDS. 2013 Mar 13;27(5):769-79 16. d' Arminio Monforte A, et al.: Insights into the reasons for discontinuation of the first highly active antiretroviral therapy (HAART) regimen in a



cohort of antiretroviral naive patients. I.CO.N.A. Study Group. Italian Cohort of Antiretroviral-Naive Patients. AIDS 2000;14:499-507.

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	Table 4. Incidence r			
	stratified by current	ate of clinical progre CD4/CD8 ratio.	ssion	
		Incidence rate per		4
	CD4/CD8 ratio	100 PYFU	95%()	
	<0.30	4.8	3.9-5.9	-
	0.30-0.45	2.4	1.9-3.1	1
	>0.45	2.0	1.7-2.3	
associ i) cu ii) cu	ation between clinical rrent CD4 cell count in rrent CD4 cell count a	on regression mod progression and the Model 1 nd current CD4/CD	els estimating	g the el 2
associ i) cu ii) cu Mode	iation between clinical rrent CD4 cell count in rrent CD4 cell count an I 1 AIC=4247	on regression mod progression and the Model 1 nd current CD4/CD ARR*	els estimating 8 in the Mode 95% Cl	g the el 2
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753

criterion (AIC) reduced by more than 4 (empiric rule from Burnham and Anderson).

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